



## Human IL-6 ELISA

Catalog Number EA-0206

(For Research Use Only)

### Introduction

Interleukin-6 (IL-6) is a multifunctional cytokine that regulates the immune response, hematopoiesis, the acute phase response, and inflammation. Deregulation of IL-6 production is implicated in the pathology of several disease processes. Its levels are observed in several diseases, including rheumatoid arthritis (RA). IL-6 plays roles in both a pro-inflammatory and anti-inflammatory cytokine. It is secreted by T cells and macrophages to stimulate immune response. In addition, like TNF $\alpha$ , IL-6 is another adipocyte secretory product that may be involved in insulin resistance. IL-6 is a cytokine secreted by many cells, including adipocytes and adipose stromal cells. IL-6 secretion is increased in the adipocytes of obese subjects (1) and may be important either as a circulating hormone or as a local regulator of insulin action (2-4). IL-6 has been implicated in the development of insulin resistance and type 2 diabetes in obese individuals. Like TNF, IL-6 inhibits the expression of LPL, but, unlike TNF, IL-6 does not stimulate lipolysis.

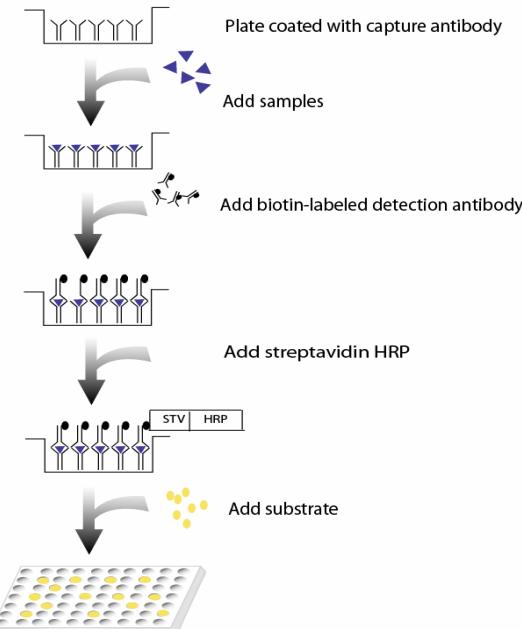


Diagram of ELISA

### Principle of the assay

IL-6 ELISA is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes an anti-human IL-6 antibody for immobilization on the microtiter wells and a biotinylated anti-human IL-6 antibody along with streptavidin conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two antibodies, resulting in the IL-6 molecules being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added, which results in the development of a blue color. The color development is then stopped with the addition of stop solution changing the color to yellow. The concentration of IL-6 is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.

### Materials provided with the kit

Component	Qty	Store at
<b>8x12 96-well microplate coated with anti-human IL-6 antibodies</b>	1	4°C
Biotin-labeled goat anti-human IL-1b antibodies	25 $\mu$ L	-20°C
Recombinant Human IL-6 standard (400ng/ml)	10 $\mu$ L	-20°C
Streptavidin-HRP conjugate	50 $\mu$ L	4°C
1X Diluent buffer	40mL	4°C
5X Assay wash buffer	40mL	4°C
Substrate	10mL	4°C
Stop solution	5mL	4°C

### Material required but not provided

- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

## Reagent preparation before starting experiment

- Dilute the 5X Assay wash buffer to 1X buffer  
40ml 5X Assay wash buffer  
160ml ddH<sub>2</sub>O
- Use serum-free conditioned media or original or 10-fold diluted sera. Sera can be diluted with 1X Diluent buffer. When serum-containing conditioned media is required, be sure to use serum as a control.
- **Standard Curve Preparation:** Prepare 8,000 pg/ml Human IL-6 standard by diluting 4 $\mu$ l of the provided Human IL-6 standard (400 ng/ml) in 200 $\mu$ l 1X Diluent Buffer. Then, do 2-fold serial dilutions six times (Standard curve is 7 wells plus 1 blank well). Add 100 $\mu$ l of the diluted standards to each well.

Standard#	IL-6 Concentration (pg/ml)
1	8,000
2	4,000
3	2,000
4	1,000
5	500
6	250
7	125
8	0

- Dilute biotin-labeled anti-human IL-6 1:400 with 1X Diluent buffer before use.
- Dilute streptavidin-HRP 1:200 with 1X Diluent buffer before use.
- Avoid contact of Substrate and Stop Solution with any metal surfaces.
- Important Note: Before beginning your experiment, quickly take 50 $\mu$ L of the Substrate and check to make sure it's clear. **If its color has changed to dark blue, do not begin the experiment and contact us immediately.**

## Assay procedure

1. Calculate the number of samples to decide how many strips need to be used.
2. Add 100 $\mu$ l of sample or standard per well and incubate for 1-2 hours at room temperature (or overnight at 4°C) with gentle shaking.
3. Aspirate each well and wash by adding 200 $\mu$ l of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Completely remove liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.
4. Add 100 $\mu$ l of diluted biotin-labeled anti-human IL-6 antibody to each well and incubate for 1 hour at room temperature with gentle shaking.
5. Repeat the aspiration/wash as in step 3.
6. Add 100  $\mu$ l of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.
7. Repeat the aspiration/wash as in step 3.
8. Before adding substrate, check to make sure it is clear. **If the substrate is already blue, please leave the wash buffer in the wells, seal and store the plate at 4°C, and contact us immediately.**
9. Add 100 $\mu$ l of substrate to each well and incubate for 10-30 minutes.
10. Add 50 $\mu$ l of Stop solution to each well. The color in the wells should change from blue to yellow.
11. Immediately measure the plate with a microplate reader at 450 nm.