



## Rat IP-10 ELISA

Catalog Number EA-3012

(For Research Use Only)

### Introduction

IFN-gamma-inducible protein (IP-10) is a member of the chemokine family of cytokines and is induced in a variety of cells in response to interferon gamma and lipopolysaccharide. It is secreted by a number of cells including monocytes, endothelial cells and fibroblasts. IP-10 plays several roles, such as chemoattraction for monocytes/macrophages, T cells, NK cells, and dendritic cells, promotion of T cell adhesion to endothelial cells, antitumor activity, and inhibition of bone marrow colony formation. Several cell types in response to IFN- $\gamma$ . IP-10 acts as potent inhibitors of angiogenesis in vivo.

### Principle of the assay

IP-10 ELISA is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes anti-rat IP-10 for immobilization on the microtiter wells and biotinylated anti-rat IP-10 antibodies along with streptavidin conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two antibodies, resulting in the IP-10 molecules being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentration of IP-10 is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.

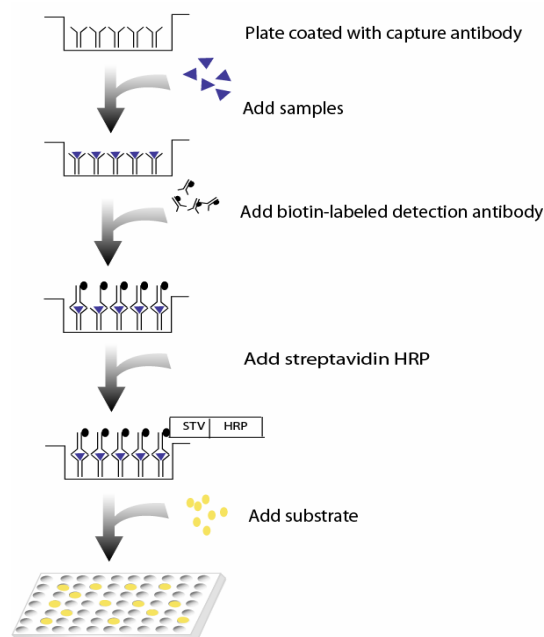


Diagram of ELISA

### Materials provided with the kit

- 8x12 96-well microplate coated with anti-rat IP-10 antibodies (4°C).
- Biotin labeled anti-rat IP-10 antibodies (-20°C).
- Streptavidin-HRP conjugate (4°C).
- Recombinant rat IP-10 standard (-20°C).
- 1X Diluent buffer (4°C).
- 5X Assay wash buffer (4°C).
- Substrate (4°C).
- Stop Solution (4°C).

### Material required but not provided

- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

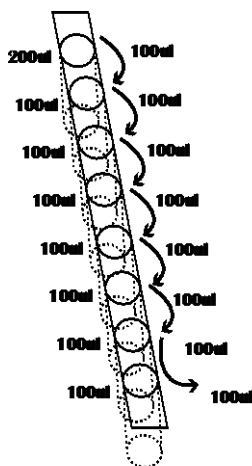
## Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer  
40ml 5x Assay wash buffer  
160ml ddH<sub>2</sub>O
- Use serum-free conditioned media or original or 10-fold diluted sera. Sera can be diluted with 1x Diluent buffer. When serum-containing conditioned media is required, be sure to use serum as a control.
- Dilute 50 times of Rat recombinant IP-10 (400ng/ml) with 1x Diluent buffer to 8ng/ml by adding 4μl Rat IP-10 recombinant protein in first well with 200μl Diluent buffer then 2-fold serial dilutions (see step 2 in Assay procedure below for the detailed instruction).
- Dilute 400 times of biotin labeled anti-rat IP-10 antibodies with 1x Diluent buffer before use.
- Dilute 200 times of streptavidin-HRP with 1x Diluent buffer before use.
- Avoid contact of Substrate and Stop Solution with any metal surfaces.
- Important Note: Before beginning your experiment, quickly take 50μL of the Substrate and check to make sure it's clear. **If its color has changed to dark blue, do not begin the experiment and contact us immediately.**

5. Add 100μl of diluted biotin-labeled anti-rat IP-10 to each well and incubate for 1 hour at room temperature with gentle shaking.
6. Repeat the aspiration/wash as in step 4
7. Add 100 μl of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.
8. Repeat the aspiration/wash as in step 4.
9. Before adding substrate, check to make sure it is clear. **If the substrate is already blue, please leave the wash buffer in the wells, seal and store the plate at 4°C, and contact us immediately.**
10. Add 100μl of substrate to each well and incubate for 10-30 minutes.
11. Add 50μl of Stop solution to each well. The color in the wells should change from blue to yellow.
12. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.

## Assay procedure

1. Calculate the number of samples to decide how many strips need to be used
2. See instruction and diagram below for standard preparation.



- a. Add 200ul 1X Diluent buffer to the 1<sup>st</sup> well. Add 100ul 1X Diluent Buffer to the rest wells of strip.
- b. Add appropriate amount of protein recombinant (follow instruction in "Reagent Preparation")
- c. Mix dilutions in 1<sup>st</sup> well and transfer 100ul from the 1<sup>st</sup> well to the next dilution. (See picture) Incubate each well for 1 hr at room temperature with gentle shaking

3. Add 100ul of sample per well and incubate for 1 hour at room temperature with gentle shaking.
4. Aspirate each well and wash by adding 200μl of 1x Assay wash buffer. Repeat the process three times for a total of three washes. Complete removal of liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.